NEW PHENOLIC ISOCULARINE AND BENZYLISOQUINOLINE ALKALOIDS AND THEIR BIOGENETIC RELATIONSHIP

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Abstract: The first three phenolic isocularines and the first diphenolic 7,8,3',4'-tetraoxygenated tetrahydrobenzylisoquinoline were isolated from Sarcocapnos crassifolia.A pathway for the biogenesis of cularine is considered.

Continuing with our search for new sources of cularine and cancentrine-type cularine (isocularine) alkaloids¹, we have now found in Sarcocapnos crassifolia the first three examples of phenolic isocularine alkaloids, namely sarcocapnidine 1, claviculine 2 and oxosarcocapnidine 3, and the first natural diphenolic 7,8,3', 4'-tetraoxygenated tetrahydrobenzyl-isoquinoline, crassifoline 4. Their structures have been established on the basis of spectroscopic data, as well as some chemical transformations and synthesis.

Crassifoline <u>4</u> was isolated as an oil, $[\alpha]_D^{=+20.6}$ (c=1.6 g/l. MeOH)². Its phenolic nature was inferred from the bathochromic shift of its UV spectrum in basic media and the presence of a broad band at 3440 cm⁻¹ in its IR spectrum. The PMR reveals the presence of two O-methyl singlets, five aromatic protons, and a methine quartet centered at δ 4.14 characteristic of an 8-oxygenated 1,2,3,4 tetrahydroisoquinoline system³. The mass spectrum showed, in addition to a molecular ion at m/e 329 (M⁺,0.1%), a base peak at m/e 192 characteristic of the tetrahydroisoquinoline molety. Finally, structure <u>4</u> for crassifoline was established by direct comparison of the alkaloid with a synthetic sample⁴.

Sarcocapnidine <u>1</u> was isolated as a white, crystalline, optically active substance, mp 126-127°C (EtOH), $\left[\alpha\right]_{D}$ =+385.4 (c=0.0696 g/l). Its molecular formula, $C_{19}H_{21}NO_4$, was obtained by elemental analysis and confirmed by MS (M⁺, base peak; f: 327,1456; c: 327,1470). The bathochromic shift of its UV spectrum in basic media together with the broad signal in its IR at 3460 cm⁻¹ revealed its phenolic nature. The PMR spectrum showed a methine proton centered at 64.48 as a doublet of doublets (dd, J_{AX} =11.8 Hz, J_{BX} =4.8 Hz) characteristic of the cularine skeleton, and four aromatic protons, two of which accidentally coincided as a singlet at $\delta 6.57$ and the remaining two as an AB quartet at $\delta_A 6.73$ and $\delta_B 6.91$ ppm J_{AB} =8.5 Hz.

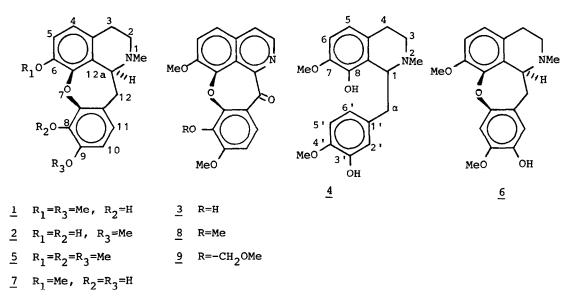
Treatment of sarcocapnidine with diazomethane yielded sarcocapnine^T 5, which proved that sarcocapnidine was a cancentrine-type cularine alkaloid. Since

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sarcocapnidine gave a positive Gibb's test⁵, the phenolic function must be located at the C-8 position. This was confirmed by the ¹³C NMR shifts⁶ observed in passing from phenol to phenoxide⁷, which established the presence of a free methine in para position to the phenolic function ($\Delta \delta = -6.38$ ppm). Structure <u>1</u> for sarcocapnidine was also supported by its synthesis from crassifoline <u>4</u> by phenolic oxidative coupling, which gave a mixture of sarcocapnidine <u>1</u> (6% yield) and O-demethylcularine <u>6</u> (3% yield)⁸.

The co-occurrence of alkaloids $\underline{1}$ and $\underline{4}$ in the same natural source and the above biogenetically patterned synthesis of $\underline{1}$ from $\underline{4}$ points to the direct oxidative coupling of a 7,8,3',4' tetraoxygenated tetrahydrobenzylisoquinoline as being the most probable among the routes proposed for cularine biogenesis^{4,8,9}.

Claviculine <u>2</u> was obtained as crystals, mp 112-1139C (EtOH) $[\alpha]_D$ =+443 (c= 0.41 g/l, MeOH), and analysed² as $C_{18}H_{19}NO_4$ (M⁺, base peak, f: 313,1323; c: 313,1309). Its phenolic nature was deduced from its UV (bathochromic shift in basic media) and IR (3420 cm⁻¹) spectra. The PMR spectrum showed a cularine-like C_{12a} methine proton centered at $\delta 4.43$ (dd, J_{AX} =11.8 Hz, J_{BX} =4.4 Hz) together with a methoxyl singlet at $\delta 3.79$ ppm. In addition, its methylation with CH_2N_2 gave sarcocapnine <u>5</u>, suggesting a diphenolic isocularine skeleton for the alkaloid. The ¹³C NMR shifts⁶ of claviculine vs its phenolate allowed us to locate the two phenolic groups at C8 ($\Delta \delta$ =-8 ppm for a methine para-carbon) and C_6 or C_9 ($\Delta \delta$ = -6.81 ppm for a quaternary para-carbon). Of the two possible structures <u>2</u> and <u>7</u> fitting in with the above structure, <u>7</u> was discarded due to the likely instability of its catechol function.



Cxosarcocapnidine <u>3</u> was isolated as yellow crystals², mp 231-2322C (MeOH), $|\alpha|_{D}=0$. It was found to contain a phenolic function (bathochromic shift on addition of base and v_{max} at 3400 cm⁻¹) and a conjugated carbonyl(1670 cm⁻¹). The

PMR showed two methoxyl singlets and three aromatic AB quartets, thus suggesting an isocularine skeleton. This was confirmed by methylation of oxosarcocapnidine with CH_2N_2 to oxosarcocapnine¹ <u>8</u>. Furthermore, the phenolic function was located at the C8 position on the basis of its partial synthesis from sarcocapnidine <u>1</u>. Thus, Fremy's salt oxidation¹⁰ of the O-methoxy-methyl protected sarcocapnidine <u>9</u> followed by acid deprotection gave <u>3</u> in 58% yield.

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- 2 All new compounds gave correct elemental analyses and the following spectroscopic data:

Crassifoline <u>4</u>:NMR(CDCl₃), δ 2.37(s, 3H, NMe), 2.43-3.16(m, 6H, 3x-CH₂), 3.84 and 3.86(ss 6H, 2x-OMe), 4.14(q, 1H, J_{AX}=8 Hz and J_{BX}=3.7 Hz, <u>H-1</u>), 4.86(broad s, 2H, -OH, disappears with D₂O), 6.57(d, 1H, J=9 Hz, <u>H</u>₆), 6.74(d, 1H, J=9 Hz, <u>H</u>₅), 6.75-6.90 (m, 3H, <u>H</u>₂, <u>H</u>₅, and <u>H</u>₆) ppm. MS m/e(%): 329(M⁺, 0.1), 193(12.5), 192(100), 177(12.5), 138(97) and 123(97).

<u>4</u> Perchlorate:mp 228-2302C(EtOH); λ_{max} (EtOH)(log ε):214(4.23), 232 sh(4.19), 282(3.82); λ_{max} (EtOH/OH⁻)(log ε):226(4.51), 246(4.18), 296(3.95).

Sarcocapnidine $1 \lambda_{max}$ (EtOH) (log ϵ):238(3.8) and 281(3.5); λ_{max} (EtOH/OH⁻) (log ϵ):250(3.73) and 294(3.62).NMR(CDCl₃) δ :2.59(s,3H,NMe), 2.74-3.5(m,6H, 3xCH₂), 4.48(dd,J_{AX}=12 Hz,J_{BX}=4.6 Hz,H_{12a}), 3.87(s,6H,2x-OCH₃), 6.57(s,2H, H₁₀ and H₁₁), 6.74(d,1H,J_{AB}=8.5 Hz,H₅) and 6.91(d,1H,J_{AB}=8.5 Hz,H₄) ppm. MS m/e(%):327.1456(C₁₉H₂₁NO₄,100%), 312(51), 310(29), 296(20), 284(24), 281(36), 174(40) and 148(14). ¹³C NMR(DMSO-d₆-dioxane as internal standard) (only non oxygenated aromatic carbons) δ (multiplicity):109.08(d), 111.12(d), 120.69(d), 121.00(s), 125.57(d), 127.32(s) and 132.10(s) ppm. ¹³C NMR(DMSO-d₆-dioxane-NaOD) δ (multiplicity):109.05(d), 110.92(d), <u>114.31(d)</u>, 120.92(s), 125.03(d), 126.66(s) and 133.00(s) ppm.

Claviculine $2:\lambda_{max}$ (EtOH) (log ε):218(4.59) and 276(4.10) nm. λ_{max} (EtOH/OH⁻) (log ε):240(5.66) and 292(4.55) nm. NMR(CDCl₃) ε :2.58(s,3H,NMe), 2.76-3.51 (m,6H,3x-CH₂-), 3.79(s,3H,-OMe), 4.47(dd,1H,J_{AX}=11.7 Hz,J_{BX}=4.5 Hz,H_{12a}), 5.03(broad s,2H,-OH,disappears with D₂O), ε .53(s,2H,ArH), ε .77(s,2H,ArH). MS m/e(ε):313.1323(M⁺,100), 298(43), 296(34), 270(13), 161(9), 148(11) and 132(16). ¹³C NMR(DMSO-d₆-D₂O) (only non oxygenated aromatic carbons), ε (multiplicity):109.48(d), 115.33(d), <u>121.62(d)</u>, 121.97(s), 126.34(d), <u>126.50(s)</u> and 131.87(s) ppm. ¹³C NMR(DMSO-d₆-NaOD) ε (multiplicity):109.50 (d), 117.85(d), <u>113.62(d)</u>, 123.04(s), 125.15(d), <u>119.69(s)</u> and 131.04(s)ppm. Oxosarcocapnidine <u>3</u>: λ_{max} (EtOH) (log ε):252(4.26), 342(3.34) and 396(3.59)nm. λ_{max} (EtOH/OH⁻) (log ε):243(4.26), 340(3.34) and 400(3.57) nm. λ_{max} (EtOH/H⁺) 2306

 $(\log \epsilon): 217(4.28), 265(4.05) \text{ and } 458(3.55) \text{ nm. NMR}(CDCl_3) \delta: 3.95(s, 3H, -O\underline{Me}), 4.98(s, 3H, -O\underline{Me}), 6.80(d, 1H, J=8.7 Hz, \underline{H}_{11}), 7.32(d, 1H, J=8.7 Hz, \underline{H}_{10}), 7.61(d, 1H, J=9 Hz, \underline{H}_5), 7.78(d, 1H, J=5.5 Hz, \underline{H}_3), 7.82(d, 1H, J=9 Hz, \underline{H}_4) \text{ and } 8.74(d, 1H, J=5.5 Hz, \underline{H}_2). MS m/e(%): 323(M, 100), 308(8), 306(14), 295(13), 280(50), 265(11), 237(16) \text{ and } 209(11).$

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